

# Package: ohtadstats (via r-universe)

March 10, 2025

**Version** 2.1.1

**Date** 2019-10-12

**Title** Tomoka Ohta D Statistics

**Description** Calculate's Tomoka Ohta's partitioning of linkage disequilibrium, deemed D-statistics, for pairs of loci. Petrowski et al. (2019) <[doi:10.5334/jors.250](https://doi.org/10.5334/jors.250)>.

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**Depends** R (>= 3.0.0)

**Imports** lattice, grDevices, stats, utils

**License** MIT + file LICENSE

**URL** <https://github.com/pfpetrowski/OhtaDStats>

**RoxygenNote** 6.1.1

**Encoding** UTF-8

**NeedsCompilation** no

**Date/Publication** 2019-11-15 11:20:02 UTC

**Additional\_repositories** <https://cranhaven.r-universe.dev>

**Repository** <https://cranhaven.r-universe.dev>

**RemoteUrl** <https://github.com/cranhaven/cranhaven.r-universe.dev>

**RemoteRef** package/ohtadstats

**RemoteSha** 9c58980697a3d9de66730d5f57910f41ffa7aa55

**RemoteSubdir** ohtadstats

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beissinger_data	<i>Chicken Genotype Data</i>
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### Description

This file is a matrix of genotypes from 96 chickens encompassing 5 breeds, genotyped as part of the Synbreed Project. Individuals are in rows. Marker genotypes are in columns, coded as 0, 1, and 2. Row names are a breed index so all rows named "1" are from breed 1, all rows named "2" are from breed 2, and so on. Column names are marker names. These data are a subset of the data used by Beissinger et al. (2016). The full dataset is hosted on Figshare at the which can be at the link below.

### Usage

```
data(beissinger_data)
```

### Format

A matrix with 1417 rows and 100 columns.

### Source

([https://figshare.com/articles/Synbreed\\_Biodiversity\\_Panel\\_Genotypes/1497961](https://figshare.com/articles/Synbreed_Biodiversity_Panel_Genotypes/1497961))

### References

Beissinger et al. (2016) Heredity. (<https://www.nature.com/articles/hdy201581>)

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dfilter	<i>Filtering datasets for subpopulations with low sample sizes</i>
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### Description

Simplifies the process of eliminating subpopulations with low sample sizes.

### Usage

```
dfilter(data, minsample)
```

**Arguments**

data	Matrix containing genotype data with individuals as rows and loci as columns. Genotypes should be coded as 0 (homozygous), 1 (heterozygous), or 2 (homozygous). Rownames must be subpopulation names and column names should be marker names.
minsamples	An integer representing the smallest number of individuals a subpopulation must contain to be included in analysis.

**Value**

filtered\_data The original dataset minus the subpopulations that fail to meet the sample size threshold.

**Examples**

```
test <- matrix(round(runif(400,1,2)), nrow = 100)
rownames(test) <- c(rep(c('A','B','C'),each=25), rep(c('D','E'), each=5), rep('F', 15))
dim(test)

#The 'D' and 'E' subpopulations have only five members each and should be removed
filtered_test <- dfilter(test,12)

dim(filtered_test) # New dataset is reduced by 10 rows (five for 'D' and five for 'E')
```

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dheatmap

*Heatmap Plot*


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**Description**

Plots a matrix of D statistics, output from dwrapper, as a heatmap.

**Usage**

```
dheatmap(d_matrix, colors = c("white", "lightblue", "blue", "darkblue",
  "black"), mode = "linear", tick.labels = TRUE, nbins = 5)
```

**Arguments**

d_matrix	A matrix of D statistics or a matrix of D statistic ratios.
colors	An optional color vector. Optionally modify the color scheme of the heatmap. If mode = 'binned', must be of length 5.
mode	A string indicating desired coloring scheme. The option "linear" scales colors linearly, "truncated" truncates values greater than 1, and "binned" returns a discretized distribution of colors.
tick.labels	A logical indicating whether or not marker labels should be drawn.
nbins	An integer specifying the number of bins to be used. Only relevant if mode is "binned".

**Details**

The `d_matrix` input should be one of the matrices output by `dwrapper`. Options are `d2it_mat`, `d2is_mat`, `d2st_mat`, `dp2st_mat`, `dp2is_mat`, `npops_mat`, `ratio1`, and `ratio2`. More customized plots can be developed using the "levelplot" package.

**Value**

A color plot

**Examples**

```
data(miyashita_langley_data)
miyashita_langley_subset <- miyashita_langley_data[,1:15]
ml_results <- dwrapper(miyashita_langley_subset)
dheatmap(ml_results[["d2it_mat"]], mode = 'linear')

## Not run:
data(miyashita_langley_data)
ml_results <- dwrapper(miyashita_langley_data)
dheatmap(ml_results[["d2it_mat"]], mode = 'linear')

## End(Not run)
```

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dparallel

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*Compute Ohta's D Statistics in a manner optimized for parallelization*


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**Description**

Infers the comparisons that this instance of the function is supposed to perform given `job_id` and `comparisons_per_job`. Returns the results of those comparisons to an SQL database.

**Usage**

```
dparallel(data_set, tot_maf = 0.1, pop_maf = 0.05, comparisons_per_job,
          job_id, outfile = "Ohta")
```

**Arguments**

<code>data_set</code>	The data set that is to be analysed.
<code>tot_maf</code>	Minimum minor allele frequency across the total population for a marker to be included in the analysis.
<code>pop_maf</code>	Minimum minor allele frequency across a subpopulation for that subpopulation to be included in analysis.
<code>comparisons_per_job</code>	The number of comparisons that each instance of <code>dparallel</code> will compute.
<code>job_id</code>	A number indicating that this is the <i>n</i> th instance of this function.
<code>outfile</code>	Prefix for the file name that results will be written to. May be a path. Do not include extension.

**Examples**

```
data(beissinger_data)
dparallel(data_set = beissinger_data,
          comparisons_per_job = 300,
          job_id = 1,
          outfile = tempfile(pattern = "beissinger_comparison", tmpdir = tempdir()))
```

dstat

*Tomoka Ohta's D Statistics***Description**

Implements Ohta's D statistics for a pair of loci. Statistics are returned in a vector in the following order: Number of populations, D2it, D2is, D2st, D'2st, D'2is.

**Usage**

```
dstat(index, data_set, tot_maf = 0.1, pop_maf = 0.05)
```

**Arguments**

index	A two-element vector of column names or numbers for which Ohta's D Statistics will be computed.
data_set	Matrix containing genotype data with individuals as rows and loci as columns. Genotypes should be coded as 0 (homozygous), 1 (heterozygous), or 2 (homozygous). Rownames must be subpopulation names and column names should be marker names.
tot_maf	Minimum minor allele frequency across the total population for a marker to be included in the analysis.
pop_maf	Minimum minor allele frequency across a subpopulation for that subpopulation to be included in analysis.

**Details**

When the loci being evaluated fail to pass the filtering thresholds determined by tot\_maf and pop\_maf, NAs are returned.

**Value**

nPops Number of subpopulations used for computation, after filtering.

D2it A measure of the correlation of alleles at two loci on the same gametes in a subpopulation relative to their expectation according to allele frequencies in the total population.

D2is Expected variance of LD for subpopulations.

D2st Expected correlation of alleles in a subpopulation relative to their expected correlation in the total population.

Dp2st Variance of LD for the total population computed over alleles only.

Dp2is Correlation of alleles at two loci on the same gamete in subpopulations relative to their expected correlation in the total population.

## References

Beissinger et al. (2016) *Heredity*. (<https://www.nature.com/articles/hdy201581>) & Ohta. (1982) *Proc. Natl. Acad. Science*. (<http://www.pnas.org/content/79/6/1940>)

## Examples

```
data(beissinger_data)
dstat(index = c(5,6), data_set = beissinger_data)
```

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dwrapper

*Ohta D Statistic Wrapper*

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## Description

Pairwise computation of Ohta's D Statistics for each pair of polymorphisms in a given dataset.

## Usage

```
dwrapper(data_set, tot_maf = 0.1, pop_maf = 0.05)
```

## Arguments

data_set	Matrix containing genotype data with individuals as rows and loci as columns. Genotypes should be coded as 0 (homozygous), 1 (heterozygous), or 2 (homozygous). Rownames must be subpopulation names and column names should be marker names.
tot_maf	Minimum minor allele frequency across the total population for a marker to be included in the analysis.
pop_maf	Minimum minor allele frequency across a subpopulation for that subpopulation to be included in analysis.

## Details

This wrapper implements the dstat function for all pairs of loci in a genotype matrix. If the input matrix includes n loci, choose(n,2) pairs are evaluated. Therefore, the computation time scales quadratically, and is not feasible for large datasets. We suggest manual parallelization across computational nodes for a large-scale (ie thousands of markers) implementation.

**Value**

A list of matrices containing the pairwise comparisons for each D statistic. Also included is the number of subpopulations evaluated in each comparison and the ratio of d2is\_mat to d2st\_mat (ratio1) and dp2st\_mat to dp2is\_mat (ratio2). The result of a comparison between marker M and marker N will be found in the Mth row at the Nth column.

**Examples**

```
data(beissinger_data)
beissinger_subset <- beissinger_data[,1:15]
dwrapper(beissinger_subset, tot_maf = 0.05, pop_maf = 0.01)

## Not run:
data(beissinger_data)
dwrapper(beissinger_data, tot_maf = 0.05, pop_maf = 0.01)

## End(Not run)
```

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miyashita\_langley\_data

*Drosophila melanogaster* genotypes

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**Description**

Genotype data obtained from Miyashita & Langley (1988). A matrix representing 85 loci in 64 individuals. Individuals are in rows. Rownames "RL", "TX", or "FK", indicate the subpopulation from which the sample was taken.

**Usage**

```
data(miyashita_langley_data)
```

**Format**

A matrix with 64 rows and 85 columns.

**References**

Miyashita & Langley (1988) Genetics 120:199-212 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1203490/>)

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